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Novel Multiple-Locus Variable-Number Tandem-Repeat Analysis Method for Rapid Molecular Typing of Human *Staphylococcus aureus*[∇]

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We evaluated the use of a novel multiple-locus variable-number tandem-repeat analysis (MLVA) method for typing of human *Staphylococcus aureus*. For a total of 150 clinical isolates, MLVA demonstrated the highest discriminatory power. MLVA correctly assigned isolates to outbreaks or identified isolates as unlinked. MLVA is a rapid and simple method for the epidemiological typing of *S. aureus*.

Pulsed-field gel electrophoresis (PFGE) is considered the gold standard method for the typing of *Staphylococcus aureus* isolates (1). Other commonly used typing schemes include multilocus sequence typing (MLST) and *spa* typing (www.mlst.net and www.spaserver.ridom.de) (2, 7). Recently, a method based on the unique lengths of the intergenic regions containing repetitive DNA loci (5, 8, 9, 12, 13), known as multiple-locus variable-number tandem-repeat analysis (MLVA), was introduced and described. Using this method, Hardy et al. described an MLVA scheme based on seven variable-number tandem repeats in *S. aureus*, termed staphylococcal interspersed repeat units (SIRUs) (5, 6). All of these methods have a number of drawbacks; most importantly, they are either fingerprinting methods, which are difficult to compare, or library typing methods lacking discriminatory power (3, 9, 10, 12, 13). We therefore developed a character-based MLVA scheme for human *S. aureus* that is based on the number of repeats of each locus, i.e., the allelic profile, which can then be used in combination with *spa* typing. A similar approach has already been reported for bovine *S. aureus* (4).

Our scheme was first tested using 100 European clinical human *S. aureus* isolates: 25 methicillin-susceptible and 75 methicillin-resistant isolates obtained between 1997 and 2004 were selected from the ENARE collection at the University Medical Centre Utrecht (UMCU), The Netherlands. These 100 specimens represented 35 MLST types and were well distributed throughout the *S. aureus* population (data not shown). MLVA typing was performed by using SIRU01, -05, -07, -13, -15, -16, and -21 (representing the *spa* gene) and *sspA* (5, 10). Each amplification was performed separately. The PCR products of SIRU01, -05, -07, -13, -15, -16, and *sspA* were analyzed on 2% agarose gel, while the SIRU21 PCR product was run on a 3% agarose gel. The number of repeat units (RUs) in each locus was determined by subtracting the sizes of the flanking regions from the size of the amplicon and then dividing the

difference by the size of the repeat (Table 1). The result was then rounded to the nearest integer value.

The MLVA showed the numbers of RUs to range from 0 to 26 (Table 1). SIRU16 and *sspA* were excluded from further study due to a lack of variation in RUs. Seventy-three isolates yielded complete MLVA profiles, and 27 isolates yielded partial profiles. The unamplified loci of the latter isolates were assigned the number 999. Combinations of RUs from SIRU01, -05, -07, -13, -15, and -21 yielded number strings that were considered to be the allelic profiles. An MLVA type (MT) was assigned to each of these profiles. The 76 MTs observed (Table 2) were clustered into three dominant MT genogroups, C1, C2, and C3 (Fig. 1). Most of the isolates belonged to clonal complexes 1, 5, 8, 12, 15, 22, 30, and 228 and were generally linked to genogroups. Isolates in clonal complex 45 were an exception. This difference may be explained either by unexpected evolutionary pressure or by the horizontal transfer of genetic material containing one or more SIRUs among the clonal complex 45 isolates.

spa typing was performed as described by Harmsen et al. (7) and yielded 50 sequence variations, or *spa* types, including 2 new *spa* types, 2285 and 2176. The numbers of repeats found in SIRU21 by *spa* typing and MLVA agreed completely.

The discriminatory power of MLVA, MLST, and *spa* typing showed that MLVA had a higher discriminatory power than both MLST and *spa* typing (for MLVA, 0.987 [95% confidence interval {CI}, 0.977 to 0.997]; for MLST, 0.941 [95% CI, 0.922 to 0.960]; and for *spa* typing, 0.963 [95% CI, 0.946 to 0.979]),

TABLE 1. Characteristics of the MLVA scheme

Locus (RU size [bp])	Formula ^a	No. of RUs (% PCR negative)
SIRU01 (55)	$(n - 157 - 30)/55$	0–5 (4.5)
SIRU05 (60)	$(n - 76 - 78)/60$	1–22 (21.5)
SIRU07 (56)	$(n - 27 - 160)/56$	1–4 (2.2)
SIRU13 (64)	$(n - 76 - 78)/64$	0–26 (2.2)
SIRU15 (131)	$(n - 48 - 174)/131$	0–5 (0)
SIRU21 (24)	$[(n - 12 - 81) - 16]/24$	1–16 (0)

^a The formula for calculating the number of RUs per locus is as follows: (amplicon size [n] – size of left flanking region – size of right flanking region)/size of RU.

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TABLE 2. MLVA allelic profiles, MTs, and *spa* types obtained from 100 *S. aureus* isolates with known MLST types

Isolate no.	Sample	MLST type	Clonal complex	<i>spa</i> type ^a	MT	No. of RUs in locus ^b					
						SIRU01	SIRU05	SIRU07	SIRU13	SIRU15	SIRU21
1	S0041	7	5	91	1	0	3	3	3	3	10
2	S0033	12	12	160	2	1	7	1	3	1	7
3	S0117	717	12	160	3	1	8	1	4	1	7
4	S0061	45	45	3	4	1	2	1	4	1	8
5	S0055	22	22	223	5	2	3	1	3	1	11
6	S0134	34	30	2285*	6	2	3	1	1	2	15
7	S0141	15	15	346	7	2	5	2	2	1	10
8	S0032	30	30	1945	8	2	3	2	1	2	12
9	S0296	36	30	18	9	2	3	2	1	2	11
10	S0084	149	5	2	10	2	2	1	4	1	10
11	S0168	228	228	1	10	2	2	1	4	1	10
12	S0072	228	228	1	10	2	2	1	4	1	10
13	S0077	228	228	1	10	2	2	1	4	1	10
14	S0085	228	228	1	10	2	2	1	4	1	10
15	S0088	228	228	1	10	2	2	1	4	1	10
16	S0065	228	228	1	10	2	2	1	4	1	10
17	S0071	736	5	1	10	2	2	1	4	1	10
18	S0056	228	228	1	10	2	2	1	4	1	10
19	S0108	228	228	41	11	2	2	1	4	1	14
20	S0074	228	228	41	11	2	2	1	4	1	14
21	S0087	228	228	1	12	2	2	1	2	1	10
22	S0062	225	5	3	13	2	2	2	3	2	8
23	S0063	228	228	1	14	2	2	2	4	1	10
24	S0070	228	228	1	15	2	2	2	1	0	10
25	S0031	5	5	3	16	2	3	1	3	1	8
26	S0054	34	30	369	17	2	3	1	2	2	11
27	S0118	225	5	3	18	2	3	1	4	1	8
28	S0075	225	5	3	18	2	3	1	4	1	8
29	S0079	225	5	3	18	2	3	1	4	1	8
30	S0081	225	5	3	18	2	3	1	4	1	8
31	S0022	714	30	136	19	2	3	1	2	2	13
32	S0047	715	30	166	20	2	3	1	2	2	12
33	S0050	30	30	122	21	2	3	2	1	0	8
34	S0057	225	5	3	22	2	3	2	3	2	8
35	S0053	30	30	18	23	2	3	2	2	2	11
36	S0021	713	30	18	24	2	4	2	2	2	11
37	S0043	26	25	81	25	2	5	2	10	5	8
38	S0049	239	8	275	26	2	3	1	8	1	8
39	S0112	5	5	2	27	2	3	1	4	1	10
40	S0128	5	5	2	27	2	3	1	4	1	10
41	S0042	228	228	1	27	2	3	1	4	1	10
42	S0045	247	8	52	27	2	3	1	4	1	10
43	S0132	739	101	56	28	2	4	2	6	2	9
44	S0111	247	8	2	29	2	1	1	4	1	10
45	S0113	36	30	12	30	2	3	2	1	2	10
46	S0060	45	45	15	31	3	2	2	1	1	10
47	S0059	5	5	2	32	3	2	3	1	1	10
48	S0027	8	8	9	32	3	2	3	1	1	10
49	S0029	8	8	8	32	3	2	3	1	1	10
50	S0044	8	8	8	32	3	2	3	1	1	10
51	S0083	738	8	8	32	3	2	3	1	1	10
52	S0068	8	8	8	33	3	3	2	2	2	10
53	S0028	239	8	37	34	3	2	2	26	1	7
54	S0120	684	8	37	35	3	2	2	1	1	7
55	S0178	239	8	30	36	3	2	2	0	1	6
56	S0177	239	8	30	36	3	2	2	0	1	6
57	S0058	225	5	3	37	4	9	2	1	1	8
58	S0030	15	15	84	38	4	7	2	2	5	11
59	S0107	239	8	37	39	4	2	2	1	1	7
60	S0109	8	8	8	40	4	2	3	0	1	10
61	S0025	247	8	51	41	4	22	2	1	1	11
62	S0035	247	8	51	42	4	4	2	0	1	11
63	S0034	15	15	254	43	4	7	2	4	5	8
64	S0297	8	8	8	44	4	1	3	0	1	10
65	S0298	8	8	64	45	4	2	2	0	1	10
66	S0127	239	8	138	46	4	2	2	0	1	6

Continued on following page

TABLE 2—Continued

Isolate no.	Sample	MLST type	Clonal complex	<i>spa</i> type ^a	MT	No. of RUs in locus ^b					
						SIRU01	SIRU05	SIRU07	SIRU13	SIRU15	SIRU21
67	S0121	685	8	64	47	4	2	3	1	1	10
68	S0129	247	8	75	48	4	4	2	1	1	12
69	S0069	239	8	30	49	5	2	2	1	1	6
70	S0066	735	45	305	50	5	3	2	4	2	11
71	S0051	239	8	421	51	2	3	1	4	1	6
72	S0052	45	45	693	52	2	2	1	4	1	1
73	S0039	101	101	56	53	2	2	1	4	1	9
74	S0067	254	8	9	74	999	2	2	1	1	10
75	S0037	9	Singleton ^c	2176*	77	4	2	999	0	1	4
76	S0300	5	5	88	80	3	10	1	999	2	11
77	S0299	72	8	126	83	3	10	999	999	2	8
78	S0295	5	5	2	85	2	999	1	4	1	10
79	S0046	228	228	1	85	2	999	1	4	1	10
80	S0076	22	22	32	87	2	999	2	4	0	16
81	S0078	22	22	32	87	2	999	2	4	0	16
82	S0089	22	22	476	89	2	999	2	2	0	11
83	S0116	97	97	527	91	2	999	3	3	1	12
84	S0024	22	22	192	93	2	999	2	7	3	13
85	S0036	45	45	1081	95	999	999	1	2	2	7
86	S0086	45	45	4	97	999	999	2	0	1	9
87	S0130	45	45	26	99	999	999	2	1	0	3
88	S0023	45	45	1933	101	999	999	2	1	1	7
89	S0166	239	8	37	103	999	999	2	0	1	7
90	S0176	239	8	37	103	999	999	2	0	1	7
91	S0167	239	8	37	105	4	999	2	0	1	7
92	S0175	239	8	37	105	4	999	2	0	1	7
93	S0026	247	8	844	107	4	999	2	0	1	8
94	S0169	247	8	51	109	4	999	2	0	1	11
95	S0110	247	8	52	111	4	999	2	0	1	10
96	S0126	572	8	51	113	4	999	2	1	1	11
97	S0119	1	1	922	115	1	999	3	1	2	6
98	S0040	188	1	189	117	1	999	3	3	5	6
99	S0115	239	8	359	119	5	999	4	3	2	9
100	S0082	737	22	5	122	2	999	2	3	0	12

^a Asterisks indicate two novel *spa* types observed in this study.

^b 999, no SIRU PCR amplification.

^c This MLST type did not group into any of the clonal complexes.

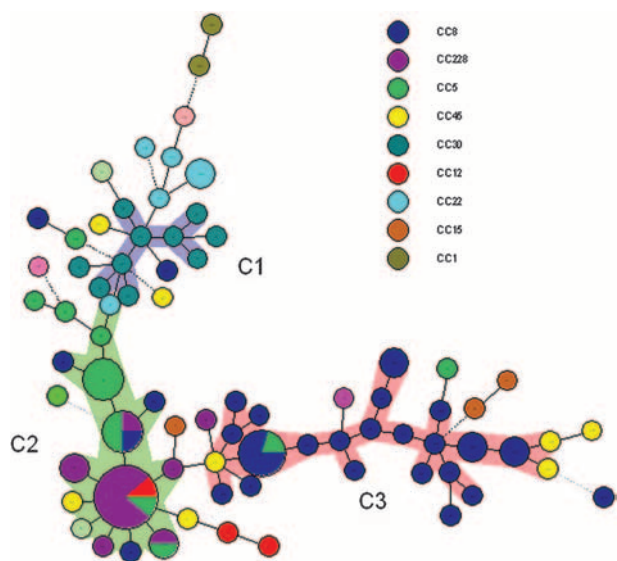


FIG. 1. Population structure of *S. aureus* isolates based on MLVA. Each circle represents a different MLVA profile. Three clusters (C1, C2, and C3) were identified as shown by the three colors (blue, green, and pink). Heavy lines connecting two MTs denote a single-locus variant, thin lines denote MTs with a double-locus variant, and dotted lines connect MTs that differ by more than two loci.

even though the 95% CI for MLVA overlapped somewhat with that for *spa* typing. The adjusted Rand index for MLVA and *spa* typing and that for MLVA and MLST were 0.341 and 0.184, respectively. The Wallace coefficients (Table 3) indicated that MLVA was reasonably predictive of both MLST and *spa* type, whereas the reverse was not true.

A confirmation study was performed to validate our MLVA scheme. This study compared the MTs with the PFGE and *spa* types of 50 *S. aureus* isolates from the Department of Infection Control and Infection Prevention of UMCU. PFGE was performed using *Sma*I as previously described by Tenover et al. (11). The isolates were considered to be either epidemiologically related or devoid of any known epidemiological link,

TABLE 3. Wallace coefficients for the methods used to characterize the 100 human *S. aureus* isolates

Typing method	Wallace coefficient for:		
	MLST	<i>spa</i> typing	MLVA
MLST		0.433	0.123
<i>spa</i> typing	0.690		0.239
MLVA	0.554	0.677	

TABLE 4. MTs, *spa* types, and PFGE profiles of *S. aureus* isolates^a

Isolate	Date of isolation	Phage type	PFGE cluster ^b	UMCU classification ^c	PFGE profile	<i>spa</i> type ^d	MT	No. of RUs in locus ^e					
								SIRU01	SIRU05	SIRU07	SIRU13	SIRU15	SIRU21
199	13 November 1997	Z136		O1	A	2175*	178	1	999	3	1	2	9
215	14 November 1997	Z136		O1	A	2175*	178	1	999	3	1	2	9
265	09 October 1998	Z136		O1	A	2175*	178	1	999	3	1	2	9
266	21 October 1998	Z136		O1	A	2175*	178	1	999	3	1	2	9
341	31 May 2000	Z136		U	F	67	89	2	1	1	4	1	9
593	01 August 2006		35	U	U	2	29	2	1	1	4	1	10
130	31 May 1996	III205		O2	B	2	29	2	1	1	4	1	10
132	10 June 1996	III205		O2	B	2	29	2	1	1	4	1	10
158	04 January 1997	III205		O2	B	2	100	2	1	1	5	1	10
241	23 June 1998	III205		U	E	447	53	2	2	1	4	1	9
478	01 March 2004		60a	U	E	447	53	2	2	1	4	1	9
298	18 June 1999	III283		U	C	2	101	2	5	1	4	1	10
319	25 November 1999	III311		U	C	2	101	2	5	1	4	1	10
519	16 December 2004		55	O3	C	2	27	2	3	1	4	1	10
524	05 January 2005		55	O3	C	2	27	2	3	1	4	1	10
530	07 January 2005		55	O3	C	2	27	2	3	1	4	1	10
529	05 January 2005		55	O3	C1	2173*	102	2	3	2	4	1	10
144	07 November 1996	Z115		O4	G	8	32	3	2	3	1	1	10
145	21 February 1996	Z115		O4	G	8	47	4	2	3	1	1	10
162	01 February 1997	Z115		O4	G	8	32	3	2	3	1	1	10
255	15 September 1998	Z115		O4	G	8	32	3	2	3	1	1	10
343	13 November 2000	Z115		U	G1	8	32	3	2	3	1	1	10
506	29 June 2004		18	U	H	8	32	3	2	3	1	1	10
516	06 December 2004		148	O5	D	8	137	3	3	3	1	1	10
518	11 December 2004		148	O5	D	8	136	3	3	3	0	1	10
520	11 December 2004		148	O5	D	8	136	3	3	3	0	1	10
347	27 March 2001	Z231		O6	I	52	412	4	25	2	1	1	10
348	27 March 2001	Z231		O6	I	52	412	4	25	2	1	1	10
363	07 April 2001	Z231		O6	I	52	412	4	25	2	1	1	10
371	13 April 2001	Z231		O6	I	52	412	4	25	2	1	1	10
395	06 December 2001	Z231		U	BB	1257	169	4	2	2	1	1	10
90	01 October 1995	III84		U	Z	138	167	4	2	2	1	1	6
125	15 February 1996	III29		U	M	31	308	999	999	2	1	0	9
129	21 May 1996	V2		U	U	1	10	2	2	1	4	1	10
284	10 November 1998	III70		U	K	24	170	4	1	3	0	1	9
331	18 February 2000	III322		U	Y	2	27	2	3	1	4	1	10
393	03 December 2001		NT	U	W	1932	90	2	3	2	2	2	10
401	05 February 2002		147	U	L	44	275	4	999	1	3	6	14
450	09 October 2003		28	U	N	728	308	999	999	2	0	1	5
459	14 November 2003		NT	U	P	567	416	3	999	3	888	2	5
466	04 December 2003		118a	U	T	311	88	2	13	1	4	1	9
482	16 June 2004		430/426	U	X	223	235	2	999	2	3	0	11
502	23 October 2004		1293	U	R	1293	179	1	999	2	2	5	8
503	28 October 2004		25a	U	S	437	190	2	999	4	2	5	7
559	13 August 2005		93	U	F2	729	237	2	999	2	3	2	10
561	26 October 2005		218	U	AA	8	168	4	1	3	1	1	10
Z548	04 March 2006		NT	U	O	11	402	3	999	3	888	2	7
570	11 March 2006		99a	U	Y	37	401	4	2	2	999	1	7
578	24 April 2006		613	U	Q	316	177	1	999	3	1	5	6
602	25 August 2006		46	U	K1	8	44	4	1	3	0	1	10

^a These classifications are compared to those previously given by the Department of Infection Control and Infection Prevention, UMCU, The Netherlands, on the basis of phage type or PFGE clustering.

^b NT, not typeable.

^c O1 to O6, outbreaks 1 to 6, respectively; U, unlinked epidemiological isolate.

^d Asterisks indicate two new *spa* types observed in this study.

^e 999, no SIRU amplification found; 888, double fragments on SIRU PCR detected.

based on either their phage or their PFGE types (depending on the method used by the National Institute of Public Health and the Environment, The Netherlands, at the time of the initial isolation) and on their times of isolation. Two PCR fragments detected for SIRU13 in two isolates were assigned the value 888.

Totals of 31 MTs, 24 *spa* types, and 30 PFGE profiles were obtained (Table 4). The Simpson indices for MLVA, *spa* typ-

ing, and PFGE were 0.971, 0.908, and 0.970, respectively. The concordance between MLVA and *spa* typing was 93.5%, and that between MLVA and PFGE was 97.7%. The adjusted Rand indices for MLVA compared with PFGE and *spa* typing were 0.599 and 0.435, respectively. The Wallace coefficients showed that MLVA and *spa* typing were mutually predictive (data not shown).

MLVA typing distinguished 10 MTs from 22 isolates belong-

TABLE 5. Evolution of *S. aureus* carriership based on MTs for four patients over time^a

Patient no.	Yr of isolation	Isolate ^b	<i>spa</i> type	MT	No. of RUs in locus ^c					
					SIRU01	SIRU05	SIRU07	SIRU13	SIRU15	SIRU21
1	1996	96-121	8	63	4	2	4	1	1	10
	1999	99-307	8	63	4	2	4	1	1	10
	2001	01-386	8	63	4	2	4	1	1	10
	2003	03-438	8	31	3*	2	2*	1	1	10
2	1999	99-288	37	35	3	2	2	1	1	7
	2001	01-346	121	56	2*	7*	2	1	1	9*
3	1987	87-A117	75	48	4	4	2	1	1	12
	1988	88-A208	75	48	4	4	2	1	1	12
	1989	89-A313	75	48	4	4	2	1	1	12
	1990	90-A358	75	48	4	4	2	1	1	12
	1992	92-A405	75	48	4	4	2	1	1	12
4	1996	96-129	1	10	2	2	1	4	1	10
	1996	96-139	1	10	2	2	1	4	1	10
	2001	01-384	1	10	2	2	1	4	1	10

^a An isolate collected in 2003 from patient 2 showed two locus variations (SIRU01 and -07) compared to the isolates collected during the previous year, even though identical *spa* types were observed. Two isolates taken from patient 3 revealed three locus variations (SIRU01, -05, and -21) indicating that these two isolates were unrelated. The different *spa* types strengthen this finding.

^b The isolates are numbered according to year of isolation and strain number, e.g., 96-121 means the isolate collected in 1996 with strain number 121.

^c Asterisks indicate locus variation.

ing to six outbreaks and 24 MTs from 28 unlinked isolates (Table 4). MLVA, *spa* type, and PFGE profile completely agreed for two of the outbreaks (O1 and O6). Four other outbreaks (O2, O3, O4, and O5) were correctly assigned when single-locus differences in MLVA, long periods between isolate collection (isolates 506, 593, 298, and 319 and isolates from O4 and O5), and a foreign origin (isolate 395) were taken into account. The 28 unrelated isolates all exhibited unique MTs.

MLVA was able to distinguish between the different outbreaks when single-locus variants were considered to belong to the same outbreak and when date of isolation was taken into consideration.

Data in Table 5 indicate that MLVA profiles are relatively stable over time. Identical MTs and *spa* types were detected in the isolates from patients 3 and 4 during a 5-year period, while variations in only two loci (SIRU01 and -07) (*spa* types remained identical) were seen in isolates from patient 1 recovered 7 years after the first isolate was taken. Distinct genotypes, MTs, and *spa* types were detected in isolates from patient 2, which may be explained by a later infection with a different *S. aureus* strain. Thus, the long-term stability of the MLVA profiles is not a major concern for outbreak detection.

In summary, our MLVA scheme showed good typeability and excellent discriminatory power for the major clonal complexes and singletons of *S. aureus*. Moreover, the reliability of the method is very good: the number of RUs determined by MLVA for SIRU21, representing the *spa* gene, completely agreed with the number of repeats obtained when *spa* typing was performed on all isolates.

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